FURANOSESQUITERPENES FROM BRACHYMERIS MONTANA*

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Abstract—Brachymeris montana afforded several furanosesquiterpenes two of them not being isolated previously. The structures were elucidated by highfield ¹H NMR spectroscopy. The present isolation of these characteristic sesquiterpenes from one of the woody South African genera of the Anthemideae is of chemotaxonomic interest.

INTRODUCTION

The small South African genus *Brachymeris* (Compositae) has not been investigated chemically. It belongs to the group of woody genera of the tribe Anthemideae, though their placement still seems to be in doubt [1]. Since, however, the chemistry of all these genera is very similar, it was of interest to see whether these compounds are also present in *Brachymeris*. We therefore have studied the constituents of *B. montana* Hutch. The results will be discussed in this paper.

RESULTS AND DISCUSSION

The roots of Brachymeris montana afforded α pinene, eugenol methyl ether, sitosterol, stigmasterol, the furanosesquiterpenes 1 [2], 3 [2], 4 [2] and 5 [2] as well as 6 and 8. The structure of 6 clearly followed from the 'H NMR spectrum, which was identical with a synthetic sample [Bornowski, H., unpublished]. The aldehyde 8, molecular formula C₁₅H₁₆O₄, showed a UV maximum at 264 nm, while the IR spectrum indicated, in addition to a band of an unsaturated aldehyde (1695 cm⁻¹), the presence of a hydroxy group and of a β -substituted furan (880 cm⁻¹). The mass spectrum showed the fragments m/z 95 and 81 typical for alkyl substituted furans as well as m/z 82, obviously formed by a McLafferty fragmentation leading to methyl furan. The ¹H NMR spectrum (Table 1) showed signals for five furan protons, one of them being drastically shifted downfield (δ 8.02), indicating that the aldehyde group was in a β -position in the furan moiety. As already followed from the UV

spectrum a further conjugation had to be assumed. Accordingly, one of the lowfield signals (δ 6.17) showed couplings with a broadened singlet at δ 4.47 (2H), the broadened triplet at 2.38 and the narrowly split double doublet at 8.02, clearly indicating the conjugation of the furan aldehyde with an olefinic

Table 1. ¹H NMR spectral data of compounds 6-8 (400 MHz, CDCl₃ TMS as int. standard)

, ,		
6	7	8
7.32 dd	5.62 d	7.38 dd
6.45 s br	3.84 dd	6.31 dd
_	2.85 ddd	_
6.25 d br		2.51 t br
}	2.56q*	
5.87 dt	-	1.84 tt
∫2.62 dddd 2.28 dddd	6.56 tq	2.38 t br
3.32 ddq	_	
		6.17 dtt
7.05 s br	6.98 s br	6.62 s br
2.08 d	2.08 d	9.91 s
7.36 s br	7.39 s br	8.02 dd
1.20 d	1.97 s br	4.47 s br
7.33 s br	_	7.25 s br
	7.32 dd 6.45 s br — 6.25 d br 5.87 dt {2.62 dddd 2.28 dddd 3.32 ddq — 7.05 s br 2.08 d 7.36 s br 1.20 d	7.32 dd 5.62 d 6.45 s br 3.84 dd - 2.85 ddd 6.25 d br 5.87 dt {2.62 dddd 2.28 dddd 3.32 ddq

^{*}Not first order.

J(Hz): compound 6: 1,2=1,15=1.5; 4,5=16; 4,6=1.3; 4,6'=1; 5,6=5,6'=6,7=6', 7=7,14=7; 6,6'=14; 12,13=1; compound 7: 1,2=2,3=2.3; 3,4=9; 3,4'=5; 5,6=7; 5,14=1.5; compound 8: 1,2=1, 15=1.7; $2,15\sim1$; 1,2=1, 15=1.7; 2,4=1; 4,5=7.5; 5,6=8; $6,8\sim1$; $6,14=8,10,13\sim1$; 8,14=1.

^{*}Part 442 in the series "Naturally Occurring Terpene Derivatives". For Part 441 see Bohlmann, F., Mathur, R., Gupta, R. K., Jakupovic, J., King, R. M. and Robinson, H. (1982) Phytochemistry 21, 2045.

The

present

double bond. As the signal at 4.47 obviously was that of an allylic CH₂OH group the sequence A was established:

Further spin decoupling showed that the remaining signals led to the sequence B, thus leading to structure 8. The stereochemistry of the 7, 8-double bond was deduced by comparison of the chemical shifts with those of similar compounds. Compound 8 we have named brachymeral. The aerial parts gave cistrans-ocimene, germacrene D, bicyclogermacrene, lupeyl acetate, eugenol methyl ether, the furanosesquiterpenes 1 [2], 2 [Bornowski, H., unpublished] and 3 as well as a further one, the epoxy lactone 7. The molecular formula of C15H16O5 immediately indicated a highly oxygenated furanosesquiterpene. From the IR spectrum followed a cross-conjugated keto group and an epoxy - γ lactone as the corresponding band was at 1815 cm⁻¹. The ¹H NMR spectrum (Table 1) was in part close to a corresponding 8 - desoxo compound which was isolated from an Ursinia species [3]. All signals and the result of spin decoupling therefore agreed nicely with the proposed structure. The relative stereochemistry at C-2 and C-3 could not be assigned with certainty. However, a cis-position of H-2 and H-3 seemed to be more likely [3]. Surprisingly, in the mass spectrum of 7, which we have named brachymerolide, the base peak was m/z 109, obviously the acyl cation formed by splitting the 7, 8-bond.

and 2 mg 7 (Et₂O-petrol, 4:1). 3 1 X = H2 2 X = 0 6 4 trans 5 cis

African genera placed in the tribe Anthemideae surely further supports that these genera, including the problematic genus *Ursinia*, are closely related. So far this type of sesquiterpene has been isolated from Athanasia [4-6], Ursinia [Bornowski, unpublished; 3, 7, 8] Lasiospermum [Bornowski, unpublished; 9], Asaemia [10], Eumorphia [2, 10], Phymaspermum [11], Stilpnophytum [10] and Gymnopentzia [11]. However, Pentzia [6, 12, 13], obviously close to Athanasia, which in part also contains compounds like *Pentzia* [6], so far only gave compounds typical for the tribe Anthemideae and from Ursinia [7, 8, 14] several sesquiterpene lactones are reported too. All these genera, however, except Pentzia, do not contain the typical acetylenes widespread in the Anthemideae. These genera therefore may be best placed in a subtribe of Anthemideae, Pentzia being a link to the other parts of the tribe.

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EXPERIMENTAL

The air-dried plant material, collected in Transvaal in February 1981 (voucher 81/76, deposited in the Botanic Research Institute, Pretoria) was extracted with Et₂Opetrol, 1:2, and the resulting extracts were separated by CC (Si gel) and further by TLC (Si gel). Known compounds were identified by comparing the ¹H NMR spectra with those of authentic material. The roots (150 g) afforded 4 mg sitosterol, 1 mg stigmasterol, 10 mg α -pinene, 5 mg eugenol methyl ether, 12 mg 1, 5 mg 3, 25 mg 4, 10 mg 5, 2 mg 6 (Et₂O-petrol, 1:3) and 6 mg 8 (Et₂O-petrol, 1:1), while the aerial parts (70 g) gave 5 mg germacrene D, 2 mg bicyclogermacrene, 3 mg lupeyl acetate, 50 mg eugenol methyl ether, 2 mg cisocimene and 2 mg trans-ocimene, 1 mg 1, 20 mg 2, 7 mg 3

Brachymerolide (7), Colourless gum, IR $\nu_{max}^{CCl_4}$, cm⁻¹: 1815 (γ -lactone), 1650 (C=CC=O); MS m/z (rel. int.); 276.100 [M]⁺

7

(16) $(C_{15}H_{16}O_5)$, 247 $[M - CHO]^+$ (3), 109 $[C_5H_5CO]^+$ (100), 81 $[109 - CO]^+$ (25).

Brachymeral (8). Colourless gum, UV (Et₂O) 264 nm, IR $\nu_{\rm max}^{\rm CCl_4}$, cm⁻¹: 3600 (OH), 2720, 1695 (CHO), 880 (β-subst. furan); MS m/z (rel. int.): 260.105 [M]⁺ (12) (C₁₅H₁₆O₄), 242 [M - H₂O]⁺ (35), 229 [M - CH₂OH]⁺ (10), 214 [242 - CO]⁺ (5), 95 [C₆H₇O]⁺ (94), 82 [C₅H₆O]⁺ (100) (McLafferty), 81 [C₅H₅O]⁺ (78).

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REFERENCES

 Heywood, V. H. and Humphries, C. J. (1977) in The Biology and Chemistry of the Compositae (Heywood, V. H., Harborne, J. B. and Turner, B. L., eds), p. 858. Academic Press, New York.

- 2. Bohlmann, F. and Zdero, C. (1978) Phytochemistry 17, 1155
- 3. Bohlmann, F. and Gupta, R. K. (1982) *Phytochemistry* 21, 1309.
- 4. Bohlmann, F. and Rao, N. (1972) Tetrahedron Letters
- 5. Bohlmann, F. and Grenz, M. (1975) Chem. Ber. 108, 357.
- 6. Bohlmann, F. and Zdero, C. (1978) Phytochemistry 17,
- 7. Bohlmann, F. and Zdero, C. (1980) Phytochemistry 19, 587.
- Bohlmann, F., Borthakur, N., Jakupovic, J. and Pickardt, J. (1981) Phytochemistry 20.
- 9. Bornowski, H. (1971) Tetrahedron 27, 4101.
- 10. Bohlmann, F. and Zdero, C. (1974) Chem. Ber. 107, 1071.
- 11. Bohlmann, F. and Zdero, C. (1972) Tetrahedron Letters
- Bohlmann, F. and Zdero, C. (1972) Tetrahedron Letters 621.
- Bohlmann, F. and Knoll, K.-H. (1979) Phytochemistry 18, 995.
- Samek, Z., Holub, M., Rychlewska, V., Grabarczyk, K. and Drozdz, B. (1979) Tetrahedron Letters 2691.